

## DIFFERENCES IN SPHINGOSINE AND FATTY ACID PATTERNS OF THE MAJOR GANGLIOSIDES OF BOVINE RETINA

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### 1. Introduction

The major ganglioside of mammalian retina is  $G_{D3}$  [1,2]. This is in contrast to other tissues of the central nervous system, in which  $G_{D3}$  constitutes only minor fractions. Furthermore, the retinal  $G_{D3}$  was found to have a fatty acid pattern different from the other major retinal gangliosides  $G_{D1a}$ ,  $G_{D1b}$  and  $G_{T1}$ , typical gangliosides of nervous tissues [2]. These findings may implicate that the  $G_{D3}$  in retina has a cellular or subcellular localization which is different from that of the other gangliosides. In this study of the gangliosides of bovine retina d18:1 sphingosine constituted more than 90% of the sphingosines in  $G_{D3}$ , while a pooled sample of  $G_{D1a}$ ,  $G_{D1b}$  and  $G_{T1}$  contained d18:1 and d20:1 sphingosines in approximately equal amounts.

### 2. Experimental

Twenty-five bovine eye bulbs were frozen immediately after slaughter. The retinæ were taken out after thawing the bulbs. Total lipids were extracted with chloroform-methanol 1:1 (v/v) and the gangliosides were isolated as previously described [2]. The  $G_{D3}$  was isolated in one batch and the  $G_{D1a}$ ,  $G_{D1b}$  and  $G_{T1}$  together in another batch. The fatty acid and sphingosine patterns of the two samples were analysed by GLC according to Vanier et al. [3].

### 3. Results

The fatty acid and sphingosine patterns of  $G_{D3}$  and of the pooled sample of  $G_{D1a}$ ,  $G_{D1b}$  and  $G_{T1}$  are presented in table 1. The predominant fatty acid was 18:0 in both samples, but  $G_{D3}$  contained less of 18:0 and more of  $C_{20}$  and  $C_{22-24}$  than the other gangliosides. A much greater difference between the two samples was recorded in the sphingosine patterns. The pooled sample of  $G_{D1a}$ ,  $G_{D1b}$  and  $G_{T1}$  contained d18:1 and d20:1 in approximately equal amounts while d18:1 constituted more than 90% of the sphingosine pattern of  $G_{D3}$ .

### 4. Discussion

Differences in fatty acid patterns of individual gangliosides from nervous tissues are reported with an increasing frequency [2-7]. According to Holm et al. [2] and Vanier et al. [3] the reported differences might be explained by a possible existence of separate sites for the biosynthesis of the individual gangliosides. This explanation was further supported by the analysis of the sphingosine patterns of the different cerebral gangliosides [3]. Thus those gangliosides which had different fatty acid patterns also differed in the sphingosine patterns.

The most significant differences in fatty acid patterns were previously obtained between  $G_{D3}$  and

Table 1  
Fatty acid and sphingosine patterns of bovine retinal gangliosides, mol-%.

Fatty acids	G <sub>D3</sub>		Sphingosines	
	G <sub>D3</sub>	G <sub>D1a</sub> +G <sub>D1b</sub> +G <sub>T1</sub>	G <sub>D3</sub>	G <sub>D1a</sub> +G <sub>D1b</sub> +G <sub>T1</sub>
18:0	64	75	d18:0	4
18:1	1	1	d18:1	91
20:0	14	12	d20:0	0
22-24	16	10	d20:1	5
				41

G<sub>D1a</sub> in mammalian retina [2]. The same differences were obtained in this study of G<sub>D3</sub> and a pooled sample of G<sub>D1a</sub>, G<sub>D1b</sub> and G<sub>T1</sub> from bovine retinae. Furthermore, between these ganglioside samples we found a difference in the sphingosine patterns to an extent never reported before. The results strongly indicate that the subcellular or cellular localization of the retinal G<sub>D3</sub> should be different from that of the retinal G<sub>D1a</sub>, G<sub>D1b</sub> and G<sub>T1</sub>.

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#### References

- [1] Handa, S. and Burton, R.M. (1969) *Lipids* 4, 205-208.
- [2] Holm, M., Månsson, J.-E., Vanier, M.-T. and Svennerholm, L. (1972) *Biochim. Biophys. Acta* 280, 356-364.
- [3] Vanier, M.-T., Holm, M., Månsson, J.-E. and Svennerholm, L. (1973) *J. Neurochem.* 21, 1375-1384.
- [4] Svennerholm, L. (1967) in: *Inborn Disorders of Sphingolipid Metabolism* (Aronson, S.M. and Volk, B.W., eds.), p. 169-186, Pergamon Press, Oxford.
- [5] Klenk, E. and Georgias, L. (1967) *Z. physiol. Chem.* 348, 1261-1267.
- [6] Ledeen, R. and Salsman, K. (1970) *Lipids* 5, 751-756.
- [7] Svennerholm, L. and Vanier, M.-T. (1972) *Adv. Exp. Med. Biol.* 19, 499-514.